

UAB MEDICAL GENOMICS LABORATORY

720 Twentieth Street South, Suite 330 Phone: (205) 934-5562
 Birmingham, Alabama 35294-0005 Fax: (205) 996-2929
www.uab.edu/medicine/genetics/medical-genomics-laboratory

UAB MGL
 Accession

For MGL Lab Use Only	Received:	Reviewed:	Accession:	Billing:	Other:
	Initials:				
	Date:				
	Comment:				

Important Notes

-This form must accompany all specimens received -All specimens received must include **two** patient identifiers and **collection date**
 -Billing information (page 5) must be included -Testing must be ordered by a qualified clinician
 Additional information is available online at www.uab.edu/medicine/genetics/medical-genomics-laboratory

Test Requisition Form

Patient Information:			Ordering Physician:				
Sample Collected: (MM/DD/YY)			<input type="checkbox"/> Please check box if physician should receive report directly				
Legal Name: (First) (MI) (Last)			Name:		NPI:		
DOB: (MM/DD/YY) MRN:			Institution:				
Address:			Address:				
City:		State:	Zip:	City:	State:	Zip:	
Sex at Birth:		SSN:	Country:		Phone:		
Parent or Guardian name (if minor):		Please check preferred result delivery:		<input type="checkbox"/> Fax:			
<input type="checkbox"/> Please check box if lab/hospital should receive report directly			<input type="checkbox"/> Email:				
Referring Lab/Hospital:			Additional Reports to:				
Name:			Name:				
Institution:			Institution:				
Address:			Address:				
City:		State:	Zip:	City:	State:	Zip:	
Country:		Phone:		Phone:			
Please check preferred result delivery:		<input type="checkbox"/> Fax:		Please check preferred result delivery:		<input type="checkbox"/> Fax:	
<input type="checkbox"/> Email:			<input type="checkbox"/> Email:				

Previous Testing History

Check all that apply:	<input type="checkbox"/> Patient or family member is pregnant. LMP:	<input type="checkbox"/> Patient has had chemotherapy in the past 6 months
	<input type="checkbox"/> Patient has had a bone marrow transplant	<input type="checkbox"/> Infectious diseases (AIDS, Hepatitis, etc.)
Has this patient or relatives had previous testing? <input type="checkbox"/> Yes <input type="checkbox"/> No		
Name/Relationship to patient:		Test/Variant/Lab:
Name/Relationship to patient:		Test/Variant/Lab:

Informed Consent

Provider's statement: I acknowledge the risks, benefits, limitations, and implications of genetic testing as outlined on the complete informed consent handout; and I have discussed the test(s) requested with the patient/guardian and I have answered his/her questions regarding testing. Informed consent has been obtained from the patient/guardian and the hard copy will be maintained.

Provider's Signature: _____



MEDICAL GENOMICS LABORATORY

720 Twentieth Street South, Suite 330
Birmingham, Alabama 35294-0005

Phone: (205) 934-5562
Fax: (205) 996-2929

UAB MGL
Accession

Name: (First) (MI) (Last)

DOB: (MM/DD/YY)

Lymphocyte/White Blood Cell-based Comprehensive Testing via Next-Gen Sequencing

RUSH Analysis: Testing completed within 15 working days of receipt of sample
(Additional \$600 RUSH fee applied; only available for tests on this page)

NF1/Legius syndrome and Other RASopathy Related Conditions

- NF1-NG: NGS and Del/Dup: *NF1* only
- NFSP-NG: NGS and Del/Dup: *NF1* and *SPRED1*
- NNP-NG: NGS: 17 genes (no *NF1*): *BRAF*, *CBL*, *HRAS*, *KRAS*, *LZTR1*, *MAP2K1*, *MAP2K2*, *NRAS*, *PPP1CB*, *PTPN11*, *RAF1*, *RASA2*, *RIT1*, *SHOC2*, *SOS1*, *SOS2*, and *SPRED1*; and Del/Dup: *SPRED1* and *LZTR1*
- RAS-NG: NGS: 18 genes: *BRAF*, *CBL*, *HRAS*, *KRAS*, *LZTR1*, *MAP2K1*, *MAP2K2*, *NF1*, *NRAS*, *PPP1CB*, *PTPN11*, *RAF1*, *RASA2*, *RIT1*, *SHOC2*, *SOS1*, *SOS2*, and *SPRED1*; and Del/Dup: *NF1*, *SPRED1*, and *LZTR1*
- CST-NG: NGS: *HRAS* only

NF2/Schwannomatosis/Meningiomatosis

- NF2-NG: NGS and Del/Dup: *NF2* only
- SCH-NG: NGS: 3 genes: *LZTR1*, *NF2*, and *SMARCB1*; and Del/Dup: *NF2*, *LZTR1*, and *SMARCB1*
- MEN-NG: NGS: 4 genes: *NF2*, *SMARCB1*, *SMARCE1*, and *SUFU*; and Del/Dup: *NF2* and *SMARCB1*

Peripheral Nerve Sheath Tumor Testing

- PNT-NG: NGS: 6 genes: *NF1*, *NF2*, *KRAS*, *LZTR1*, *PTPN11* and *SMARCB1*; and Del/Dup: *NF1*, *NF2*, *LZTR1*, and *SMARCB1*

McCune-Albright Syndrome

- GNAS-NG: NGS: *GNAS* exons 8 and 9 only

Rhabdoid Tumor Predisposition Syndrome

- RT-NG: NGS: *SMARCB1* and *SMARCA4*; and Del/Dup: *SMARCB1* only

Tuberous Sclerosis Complex

- TSCP-NG: NGS and Del/Dup: *TSC1* and *TSC2*

Capillary Malformation Arteriovenous Malformation Syndrome

- RASA-NG: NGS: and Del/Dup: *RASA1* and *EPHB4*

Additional Information

Test Description Key:

Next Generation Sequencing (NGS)
Deletion/Duplication analysis (Del/Dup)

For additional testing options via tumor/biopsy, please see page 3 of this order form.
Please contact the lab via phone (205) 934-5562 or via email at medgenomics@uabmc.edu if you have any questions when completing this form.
For additional information, visit our website at www.uab.edu/medicine/genetics/medical-genomics-laboratory

Important points of consideration for testing

The average coverage for all of our panels is >1600x. Specifically for the *NF1* gene, the NGS approach covers >98% of the *NF1* coding region at ≥350X and 99% ≥200X, allowing detection of very low level mosaicism, down to 3-5% variant allele fraction respectively. For all other genes on our panels, the NGS approach covers an average of 99% at ≥200X. Remaining regions are covered at ≥30X. However, for patients with segmental/mosaic presentation, deep coverage in lymphocyte cells may be insufficient to identify the underlying gene change. Testing the affected tissue(s) may be necessary to confirm a diagnosis. Please see page 3 for our tumor/biopsy-based testing options.

Please note: For patients with an ongoing pregnancy who require comprehensive NF1 testing, "NF1-R" is recommended due to the sensitivity and fast turnaround time of this test (please see page 4 for this option).

Specimen Requirements

Accepted Specimens

Specimen requirements vary based on test requested; please see our website for more details.

- Blood: 3-6ml EDTA (receipt within one week of collection)
- Saliva: OGR-575 DNA Genotek (kits are provided upon request)
- DNA: extracted from lymphocyte cells, a minimum of 25ul at 3µg, O.D. value at 260:280nm ≥1.6 (must be extracted in a CLIA or equivalent certified lab)
- Fibroblast cells

Specimen Information:

- Peripheral Blood (EDTA); # Tubes: _____
- Extracted DNA; Source: _____
- Saliva (kit must be provided by MGL)
- Other, please describe: _____

Please note: failure to provide a date of collection can delay release of results

Sample Collected Date (required): _____

Name: (First) (MI) (Last) DOB: (MM/DD/YY)

Tumor/Biopsy-based Comprehensive Testing

Please check here if blood or DNA is provided for confirmation testing. Blood Collected: (MM/DD/YY)

NF1/SPRED1 on biopsied CALs and Neurofibromas

****Please contact the laboratory at least one week in advance of the biopsy before ordering this test as media must be provided in advance and special shipping instructions apply. Biopsies must arrive within 60 hours of collection****

- NF14C: Sanger(RNA) and Del/Dup: *NF1* (with automatic reflex to *SPRED1*) on biopsied CALs
- NF14N: Sanger(RNA) and Del/Dup: *NF1* on biopsied neurofibromas

NF2-, LZTR1-, SMARCB1-related Schwannomatosis

- NF2-NG: Fresh/Frozen Tumor or Tumor Block for NGS with reflex to Sanger as needed and Del/Dup: *NF2* only
- SCH-NG: Fresh/Frozen Tumor or Tumor Block for NGS with reflex to Sanger as needed and Del/Dup: *NF2, LZTR1, and SMARCB1*

Rhabdoid Tumor Predisposition Syndrome

- RT-NG: Fresh/Frozen Tumor or Tumor Block for NGS with reflex to Sanger as needed *SMARCB1* and *SMARCA4*; and Del/Dup: *SMARCB1* only

RASopathy Related Conditions

- NNP-NG: Fresh/Frozen Tumor for NGS (no *NF1*) or Tumor Block for NGS with reflex to Sanger as needed: *BRAF, CBL, HRAS, KRAS, LZTR1, MAP2K1, MAP2K2, NRAS, PPP1CB, PTPN11, RAF1, RASA2, RIT1, SHOC2, SOS1, SOS2, and SPRED1*; and Del/Dup: *SPRED1* and *LZTR1*
- RAS-NG: Fresh/Frozen Tumor or Tumor Block for NGS with reflex to Sanger as needed: *BRAF, CBL, HRAS, KRAS, LZTR1, MAP2K1, MAP2K2, NF1, NRAS, PPP1CB, PTPN11, RAF1, RASA2, RIT1, SHOC2, SOS1, SOS2, and SPRED1*; and Del/Dup: *NF1, SPRED1, and LZTR1*

Meningiomatosis

- MEN-NG: Fresh/Frozen Tumor or Tumor Block for NGS with reflex to Sanger as needed: *NF2, SMARCB1, SMARCE1, and SUFU*; and Del/Dup: *NF2* and *SMARCB1*

Peripheral Nerve Sheath Tumor Testing

- PNT-NG: Fresh/Frozen Tumor for NGS with reflex to Sanger as needed: *NF1, NF2, KRAS, LZTR1, PTPN11* and *SMARCB1*; and Del/Dup: *NF1, NF2, LZTR1, and SMARCB1*

Tuberous Sclerosis Complex

- TSC-NG: Fresh/Frozen Tumor or Tumor Block for NGS with reflex to Sanger as needed and Del/Dup: *TSC1* and *TSC2*

Additional Information

Test Description Key:

Next Generation Sequencing (NGS)
 Sanger Sequencing (Sanger)
 Deletion/Duplication analysis (Del/Dup)

Please contact the lab via phone (205) 934-5562 or via email at medgenomics@uabmc.edu if you have any questions when completing this form.
 For additional information, visit our website at www.uab.edu/medicine/genetics/medical-genomics-laboratory

Important points of consideration for testing

- The MGL offers next generation sequencing testing options that provide the ability to identify variants (indels and substitutions) as low as 3% of the alleles, depending on coverage in the regions of interest.
- NF1/SPRED1 biopsy-based testing is considered the "gold standard" approach for confirming a diagnosis of mosaic/segmental NF1 or Legius Syndrome.
- A minimum of two biopsies is required for NF1 testing. Two or more tumors are suggested for our other testing options. There are no additional fees associated with testing on additional biopsy specimens.
- When proceeding with tumor-based testing for NF2-related SWN, test code "SCH-NG" (*NF2, SMARCB1, and LZTR1*) is suggested unless the patient has findings unique to NF2.

Specimen Requirements

Accepted Specimens

Specimen requirements vary based on test requested; please see our website for more details.

- CALs or Neurofibromas: require special media transport (kits are provided upon request, to be arranged at least one week in advance of procedure)
- Fresh/Frozen Tumors: please submit a pathology report; for additional requirements, see tumor submission checklist
- Formalin-Fixed Paraffin-Embedded Tumors (Tumor Block): please submit a pathology report; blocks are preferred to curls, when available; for additional requirements, see tumor submission checklist

Specimen Information:

- Frozen Fresh
- Paraffin Curls Paraffin Block
- Extracted DNA; Source: _____
- Biopsy-CAL-spot; # biopsies: _____
- Biopsy-Neurofibroma; # biopsies: _____
- Please note: failure to provide a date of collection can delay release of results
- Tumor Collection Date (required): _____

Name: (First) (MI) (Last) DOB: (MM/DD/YY)

Sanger Testing from Blood/Saliva/DNA

<p>NF1/Legius syndrome and Other RASopathy Related Conditions</p> <p><input type="checkbox"/> NF1-R: Sanger and Del/Dup: <i>NF1 (RNA)</i></p> <p><input type="checkbox"/> NFSP-R: Sanger and Del/Dup: <i>NF1 (RNA)</i> and <i>SPRED1 (gDNA)</i></p> <p align="center">Von Hippel-Lindau</p> <p><input type="checkbox"/> VHL1: Sanger and Del/Dup: <i>VHL</i></p> <p align="center">PTEN-Related Disorders</p> <p><input type="checkbox"/> PTEN1: Sanger and Del/Dup: <i>PTEN</i></p> <p align="center">Fragile X syndrome</p> <p><input type="checkbox"/> FRX: PCR and, if needed, reflexive confirmatory testing by Southern blot analysis: <i>FMR1</i></p>	<p>Medium Chain Acyl-CoA Dehydrogenase Deficiency (MCADD)</p> <p><input type="checkbox"/> MCD1: Targeted analysis of exon 11 and, if needed, reflexive full gene sequencing by Sanger: <i>ACADM</i></p> <p align="center">Autosomal Recessive Polycystic Kidney Disease</p> <p><input type="checkbox"/> PKDL: Linkage Analysis for informativity</p> <p><input type="checkbox"/> PKDPL: Prenatal Linkage (see Prenatal Specimen Requirements)</p> <p>FATHER: <u>Name and DOB (mm/dd/yyyy)</u></p> <p>MOTHER: <u>Name and DOB (mm/dd/yyyy)</u></p>
--	---

Known Variant Testing

KT2: Targeted detection of a specific, previously identified known variant in any gene that is available at our lab by Sanger sequence, MLPA, and/or FISH analysis (Complete Previous Testing History: Page 1)

KT2-NG: Targeted testing for a known variant with deep coverage of the alleles and detection of mosaicism for a variant present in at least 3% of alleles (Complete Previous Testing History: Page 1)

RT2: Targeted RNA-based testing for VOUS found during Next Generation Sequencing (Complete Previous Testing History: Page 1)

PT2: Prenatal testing (see Prenatal Specimen Requirements; Complete Previous Testing History: Page 1)

MCC: Blood specimen for mother provided for maternal cell contamination studies (required if not previously tested)

Other (unlisted options, please indicate below)

Please contact lab before selecting this option

Additional Information

<p><u>Test Description Key:</u> Next Generation Sequencing (NG) Sanger Sequencing (Sanger) Deletion/Duplication analysis (Del/Dup)</p>	<p>Please contact the lab via phone (205) 934-5562 or via email at medgenomics@uabmc.edu if you have any questions when completing this form. For additional information, visit our website at www.uab.edu/medicine/genetics/medical-genomics-laboratory</p>
---	---

Specimen Requirements

<p align="center">Accepted Prenatal Specimens</p> <p>Specimen requirements vary based on test requested; please see our website for more details.</p> <p>-Direct CVS: minimum 10 mg cleaned villi</p> <p>-Direct amniotic fluid: minimum 10 ml fluid</p> <p>-Cultured CVS: Two T25 flasks (>70% confluent)</p> <p>-Cultured amniocytes: Two T25 flasks (>70% confluent)</p>	<p align="center">Prenatal Specimen Information:</p> <p><input type="checkbox"/> Amniotic fluid <input type="checkbox"/> Direct CVS (cleaned)</p> <p><input type="checkbox"/> Cultured amniocytes <input type="checkbox"/> Cultured villus cells</p> <p>Location of back-up culture (required):</p> <p>Sample Collected Date (required):</p>
<p align="center">Accepted Specimens</p> <p>Specimen requirements vary based on test requested; please see our website for more details.</p> <p>-Blood: 3-6ml EDTA (must arrive within 60-72 hours of collection for RNA-based tests)</p> <p>-DNA: extracted from lymphocyte cells, a minimum of 25ul at 3µg, O.D. value at 260:280nm ≥1.6 (must be extracted in a CLIA or equivalent certified lab)</p> <p>-Sperm (for KT2-NG only): Fresh, sterile semen collection using a local sperm bank/cryobank facility</p>	<p align="center">Specimen Information:</p> <p><input type="checkbox"/> Peripheral Blood (EDTA); # Tubes:</p> <p><input type="checkbox"/> Extracted DNA; Source:</p> <p><input type="checkbox"/> Other, please describe: _____</p> <p><u>Please note: failure to provide a date of collection can delay release of results</u></p> <p>Sample Collected Date (required):</p>

UAB MEDICAL GENOMICS LABORATORY

720 Twentieth Street South, Suite 330 Phone: (205) 934-5562
 Birmingham, Alabama 35294-0005 Fax: (205) 996-2929
www.uab.edu/medicine/genetics/medical-genomics-laboratory

UAB MGL
 Accession

Name: (First) (MI) (Last)	DOB: (MM/DD/YY)
---------------------------	-----------------

Billing

Please hold sample until further notice from the ordering facility.

Important Information

By completing this form, you agree that you have discussed the MGL's billing policies with your patient.

Credit card information **MUST** be provided with sample submission for self-pay clients. Please note: If you are paying via self-payment or requesting a benefits investigation, there will be a 3-5 working day delay on the initiation of your test. Requests for cancellation, test change, or billing method change of ongoing testing must be submitted to the laboratory within three working days of specimen arrival. Individuals or institutions submitting requests after the three working day window may still incur charges for the cost of testing.

Full information on the billing policies is available at www.uab.edu/medicine/genetics/medical-genomics-laboratory

 Institutional Bill

Please check box if billing institution should receive report directly

Institution:	PO# (if applicable):
--------------	----------------------

Address:

City:	State:	Zip:
-------	--------	------

Contact (Name and Title):	Preferred method of contact: <input type="checkbox"/> Email <input type="checkbox"/> Phone
---------------------------	---

Email:	Phone:	Fax:
--------	--------	------

 Self-Payment Enclosed *PLEASE ENSURE ALL INFORMATION IS LEGIBLE*

Visa MasterCard Discover American Express

Name as it appears on card:

Card Number:	Expiration: (MM/YY)	3-digit Security Code:
--------------	---------------------	------------------------

Cardholder's Signature:	Preferred method of contact: <input type="checkbox"/> Email <input type="checkbox"/> Phone
-------------------------	---

Email:	Phone:
--------	--------

 Bill Third Party Insurance Company

Please include a copy of the pre-approval statement or provide the approval number if payment has been pre-authorized in advance of shipment.

Insurance Carrier: _____
 Insurance pre-verification/authorization previously performed? Yes No If yes, approval number is required: _____

Please check box if you would not like insurance pre-verification/authorization to be performed by the MGL.

Please send a legible copy of the patient's insurance card, front and back.

ICD-10 Codes (required):

Important Considerations for Insurance Billing

For a list of contracted insurance companies, please visit our website or call our billing coordinator at 205-934-5523. As insurance prices are not listed, please call the billing coordinator to request a quote, if needed.

The MGL will contact the insurance provider to inquire regarding the CPT code coverage for all samples submitted for insurance payment. The healthcare provider will be contacted with the copay/deductible and also in cases where the insurance provider denies coverage of the requested codes or supporting documents are required from the provider to confirm coverage. The ordering provider/clinician's office is responsible for obtaining prior authorization, if it is required. This service is not offered for prenatal samples.

Please note: An insurance verification is not a guarantee of payment. Out of State Medicaid is not accepted under any circumstances. All RUSH fees must be paid up front. By completing this form, you agree that you have discussed the MGL's billing policies with your patient.

Informed Consent for Genetic Testing

This form does not need to be returned to the Medical Genomics Laboratory if Informed Consent portion of the Test Request form has been signed.

I hereby consent for:

Name:	DOB:	Gender:
-------	------	---------

To participate in genetic testing for the following RNA/DNA-based cascade of tests ordered by my physician at the University of Alabama at Birmingham (UAB) Medical Genomics Laboratory (MGL):

Genetic Tests:

I understand that:

1. Any biological samples submitted for genetic testing (e.g., blood, cheek cells, saliva, amniotic fluid, chorionic villi, tumor, and/or tissue) will be removed from me and/or my minor child(ren) using standard techniques which carry their associated risks.

2. Any samples obtained will be used for the purpose of attempting to determine if I and/or members of my family carry genetic changes in the disease genes ordered by my physician.

3. The genetic tests performed at the MGL are the most sensitive developed and are highly specific. However, sensitivity and specificity are test-dependent. Additional testing details and the specific detection rates of each test can be found at www.genetics.uab.edu/medgenomics.

4. The following are possible outcomes for the specific tests listed above:

Positive	Unknown Significance	Negative
This is an indication that I may be predisposed to or have the specific disease, or condition tested. Further testing may be needed to confirm the diagnosis.	There may be a possibility that the laboratory findings will be ambiguous or of unknown significance. This may require additional testing from me or my family members. In rare circumstances, findings may be suggestive of a condition different than the diagnosis that was originally made.	There is a chance that I will still have this genetic condition even though the genetic test results are negative. Due to limitations in technology and incomplete knowledge of genes, some changes in RNA/DNA or protein products that cause disease may not be detected by this test.

5. In other cases, the RNA/DNA test is unable to identify an abnormality although the abnormality may still exist. This event may be due to incomplete knowledge of the gene structure or an inability of current technology to identify certain types of mutations in the gene. When clinically necessary, the MGL may use a method called linkage analysis. This method is not a direct test, but will report the probability that you and/or family members have an inherited disease or disorder. In some families, the markers used in linkage analysis may be uninformative. If so, linkage testing cannot provide results for the family members in question.

6. The RNA/DNA analysis performed by the MGL is specific for the genetic test listed above and in no way guarantees my health or the health of my living or unborn children. The MGL cannot be responsible for an erroneous clinic diagnosis made elsewhere.

7. The tests performed at the MGL are expanded and improved continuously. The tests offered are not considered research but are considered the best and newest laboratory service that can be offered. Genetic testing is complex and utilizes specialized materials so there is always some very small possibility that the test will not work properly or that an error will occur. There is a low error rate (perhaps 1 in 1000 samples) even in the best laboratories. Additionally, in very rare instances, this test may reveal an important genetic change that is not directly related to the clinical reason for ordering this test. This would be considered an incidental finding. The MGL reserves the right to report these incidental findings if they are clinically relevant to the patients and/or their families. In such instances, these results will be discussed with my healthcare provider and additional testing may be recommended. My signature acknowledges my voluntary participation in this test, but in no way releases the laboratory and staff from the MGL from their professional or ethical responsibility to me.

8. The MGL does not return DNA samples to individuals or physicians. While the MGL is not a specimen banking facility, in some cases it may be possible for the laboratory to reanalyze my remaining DNA upon request. The request for additional studies must be ordered by my referring physician/counselor and there will be an additional fee.



MEDICAL GENOMICS LABORATORY: NF2 & SCHWANNOMATOSIS PHENOTYPIC CHECKLIST FORM



Patient ID: _____

Referring Physician: _____ Date of Exam ___/___/___

DEMOGRAPHIC INFORMATION

Gender: Male Female Date of Birth: ___/___/___

Ethnicity: Mother: White Black Native American Hispanic Asian Other:
Father: White Black Native American Hispanic Asian Other:

DIAGNOSIS

Does the patient have a clinical diagnosis of NF2? Yes No If Yes, age at diagnosis: _____

Family history: Sporadic Familial Unknown

SIGNS AND SYMPTOMS

Ear: Absent Unknown Present: Bilateral Deafness Unilateral Deafness Age of symptoms: _____
 Balance Dysfunction Tinnitus
 Audiometric Abnormality, please describe: _____
 Other, please describe: _____

Eyes Absent Unknown Present: Blindness Age of Symptoms: _____
 Lenticular opacities
 Lisch nodules
 Left Right
 Bilateral Unknown

Schwannomas

Vestibular schwannomas Age of Symptoms: _____
 Bilateral
 Unilateral
 No evidence of vestibular nerve tumor by MRI (Age: ___ yrs)
 Lack of symptoms of vestibular nerve tumor, but no MRI done (Age: ___ yrs)
 Unknown

Spinal schwannomas Age of Symptoms: _____
 Present
 No evidence by MRI (Age: ___ yrs)
 Lack of symptoms, but no MRI done (Age: ___ yrs)
 Unknown
Provide location of spinal tumors: C ___ to C ___, T ___ to T ___, L ___ to L ___

Other schwannomas Age of Symptoms: _____
 Absent Unknown
 Head Neck Trunk L Arm L Hand L Leg L Foot
 Abdomen Pelvis Genital area R Arm R Hand R Leg R Foot

Present only in an anatomically limited distribution(single limb or segment of the spine): yes or no

Result *SMARCB1*-staining on the tumor specimen: Abnormal (no *SMARCB1*-staining)
 Normal (*SMARCB1* staining)
 Not performed



MEDICAL GENOMICS LABORATORY: NF2 & SCHWANNOMATOSIS PHENOTYPIC CHECKLIST FORM



Meningiomas

- Present, Location: _____
- No evidence by MRI (Age: ___yrs)
- Unknown

Age of Symptoms: _____

Other spinal tumors

- Absent by MRI
 - Present, asymptomatic
 - Present, symptomatic
 - Unknown
- Pathology Known: Yes, please specify: _____ No

Age of Symptoms: _____

Provide location of spinal tumors: C ___ to C ___, T ___ to T ___, L ___ to L ___

Cranial nerve involvement

- Present, Location: _____
- No evidence by MRI (Age: ___yrs)
- Unknown

Age of Symptoms: _____

Skin

CAL spots

- 1
- 2-3
- 4-5
- >5-10
- >10

Age of Symptoms: _____

Neurofibromas

- 1-5
- ≥6-99
- ≥100

Age of Symptoms: _____

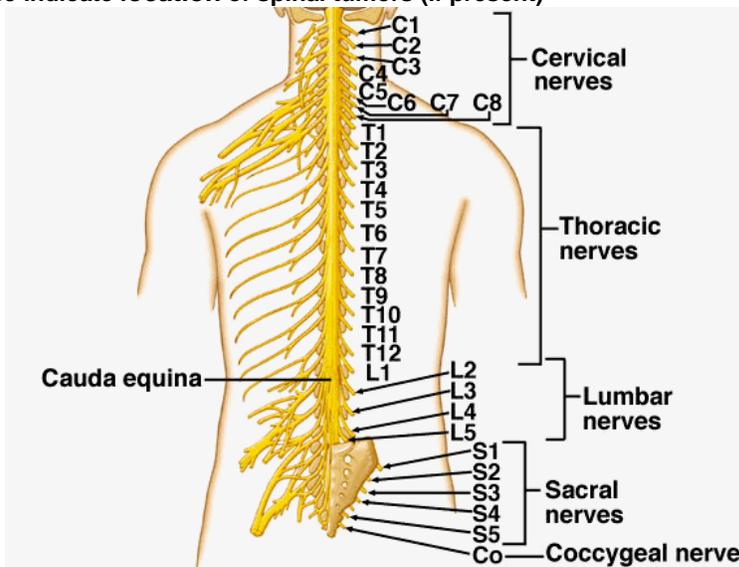
Skin fold freckling

- | | Left | Right |
|------------|--------------------------|--------------------------|
| Groin | <input type="checkbox"/> | <input type="checkbox"/> |
| Axilla | <input type="checkbox"/> | <input type="checkbox"/> |
| Submammary | <input type="checkbox"/> | <input type="checkbox"/> |

Age of Symptoms: _____

Please indicate location of spinal tumors (if present)

Additional comments/remarks:



Tumor Specimen Submission Checklist

The following requirements must be met in order to process tumor specimens. The UAB Medical Genomics Lab now proudly offers Fresh/Frozen/FFPE Tumor testing utilizing Next Generation Sequencing. Please confirm that the submitted tissue meets each requirement by placing a check mark next to each statement. If your pathology department is unable to confirm this information for you, please select the check-box below. If Neurofibromatosis Type 1 is your primary concern, neurofibromas and biopsied CALs still require being collected in our media and are run via Sanger sequencing. Please contact the MGL at (205) 934-5562 to request collection media for these sample types.

For NGS Sequencing

Fresh Tumor Specimen Checklist	
	This tumor is at least 5mm-cubed
	This specimen contains at least 60% pure tumor content
	Each specimen has been sent in individual vials of basic, sterile culture media such as RPMI or PBS and is marked with its specific location and/or tumor type
Frozen Tumor Specimen Checklist	
	This tumor is at least 5mm-cubed
	This specimen contains at least 60% pure tumor content
	This specimen has been snap frozen and sent on dry ice and is marked with its specific location and/or tumor type

Formalin-fixed paraffin embedded block	
	This tumor block has a surface area of a least 5mm squared <u>or</u> This specimen contains at least 3-6 loose paraffin curls (no slides) that are 30-50 microns thick
	This tumor specimen contains greater than 70% nucleated cells
	This specimen contains at least 60% pure tumor content
Notes or Special Comments	